

Morphological variation on isolated populations of *Praocis* (*Praocis*) spinolai

Hugo A. Benítez^{1,2a*}, Jaime Pizarro-Araya^{3b}, Raffaella Bravi^{4c}, María-José Sanzana^{5d}, Fermín M. Alfaro^{3,6e}

Faculty of Life Sciences, University of Manchester, Michael Smith Building, Oxford Road, Manchester M13 9PT, UK

²Instituto de Alta Investigación, Universidad de Tarapacá, Casilla 7-D Arica, Chile

³Laboratorio de Entomología Ecológica, Departamento de Biología, Universidad de La Serena, Casilla 599, La Serena, Chile

⁴Environmental Biology Department, University Roma Tre, V. le G. Marconi 446, 00146, Rome, Italy

⁵Departamento de Zoología, Facultad de Ciencias Naturales y Oceanográficas, Universidad de Concepción, Concepción, Casilla: 160-C, Chile

⁶Laboratorio de Genética y Evolución, Departamento de Ciencias Ecológicas, Facultad de Ciencias, Universidad de Chile, Santiago, Chile

Abstract

In this study, the morphological variations of four geographically isolated populations of *Praocis* (*Praocis*) spinolai Gay & Solier (Coleoptera: Tenebrionidae) in the transitional coastal desert, Chile, were studied. The study was conducted in the coastal area of Punta de Choros and Los Choros-Archipelago, which includes three islands: Choros, Damas, and Gaviota. 113 specimens of the species *P. (P.) spinolai* belonging to the four locations sampled were collected analyzed with geometric morphometrics techniques to explore the pattern of shape variation on the different isolated environments. The principal component analysis revealed a well-defined pattern of variation between the populations analyzed. Moreover, differences between populations emerged also from the canonical variation analysis and were confirmed by the Procrustes ANOVA. All analyses performed confirmed the existence of a pattern of variation, due to the isolation of the populations and to environmental effects. The islands are subject to more arid pressures than the continent, where there is a more stable environment and the presence of coastal wetlands and the coastal range of mountains act together and enable fog condensation. This study indicates the existence of a clear pattern of variation, which indicates an evolutionary trend among the population examined.

Keywords: coastal desert, epigean tenenebrionids, geometric morphometrics, Pingüino de Humboldt National Reserve **Correspondence:** a hugo.benitez@postgrad.manchester.ac.uk, b japizarro@userena.cl, c raffaella.bravi@uniroma3.it, d

mariajosanzana@udec.cl, e fermin.alfaro@ug.uchile.cl, *Corresponding author

Editor: Felipe Soto was editor of this paper.

Received: 11 June 2013 Accepted: 6 November 2013 Published: 26 January 2014

Copyright: This is an open access paper. We use the Creative Commons Attribution 3.0 license that permits unrestricted use, provided that the paper is properly attributed.

ISSN: 1536-2442 | Vol. 14, Number 11

Cite this paper as:

Benítez HA, Pizarro-Araya J, Bravi R, Sanzana M-J, Alfaro FM. 2014. Morphological variation on isolated populations of *Praocis* (*Praocis*) spinolai. Journal of Insect Science 14:11. Available online: http://www.insectscience.org/14.11

Introduction

Over the last century, research on islands has continued to advance the understanding of the evolutionary process (Emerson 2008). Island archipelagos provide unique scenarios for studying the roles of geography and ecology in driving population divergence and speciation while playing a crucial role in the diversification of biotas. Oceanic islands also have long been recognized as natural laboratories for the study of evolutionary processes (Mayr 1967; Grant 1998; Losos and Ricklefs 2009; Mila et al. 2010).

The Pingüino de Humboldt National Reserve is located on the coastal border between Huasco (Atacama Region) and Elqui (Coquimbo Region) provinces in Chile, and it comprises a total area of 859.3 ha. It was created in 1990 and is a part of the country's National System of Protected Wild Areas. A portion of the reserve encompasses the Choros Archipelago, which includes the islands of Choros, Damas, and Gaviota. These islands are located on the northwestern end of the Punta Choros area, Coquimbo Region (Castro and Brignardello 2005), and constitute a peculiar insular ecosystem. The three islands are located in an area within the transitional coastal desert (25–32° Lat S), the latter of which is characterized by the presence of an unusually-species-rich arthropod fauna (Cepeda-Pizarro et al. 2005; Valdivia et al. 2008, 2011; Pizarro-Araya et al. 2008; Alfaro et al. 2009; Alcayaga et al. 2013), endemism (Jerez 2000; Pizarro-Araya and Flores 2004; Ojanguren-Affilastro et al. 2007: Pizarro-Arava et al. 2012a.b: Oianguren-Affilastro and Pizarro-Araya in press; Laborda et al. 2013), and restricted distribution (Pizarro-Araya and Jerez 2004; Agusto et al. 2006; Alfaro et al. 2013).

It is well known that adaption over time to a specific environment is the result of both environmental pressures and geographic distances. affecting geographic environments at a local scale and thus their associated flora and fauna (Alibert et al. 2001; Cepeda-Pizarro et al. 2003; Benítez et al. 2008; Benítez 2013). Moreover, it is well documented that adverse temperatures, nutripresence of chemicals, tional stress, population density, and many other factors that cause stress during development can lead to an increase in the presence of morphological asymmetries as a result of high intraspecific variation (e.g., Rettig et al. 1997; Benítez et al. 2008; Benítez 2013). Therefore, it is expected that when environmental conditions change, organisms and populations should adapt to the new conditions (Clarke 1993). In this context, adaptive variation plays a major role because it reflects historical evolution and determines the population's phenotypic response. Floate and Fox (2000) and Piscart et al. (2005) suggested that the degrees of phenotypic disturbance reflect the ability of an individual to overcome the effects of stress. In fact, in epigean arthropods the more symmetrical individuals would have a greater survival chance than those with any level of asymmetries.

Among epigean arthropods, Tenebrionidae (Coleoptera) constitute a characteristic group of the arid and semiarid ecosystems fauna (Cloudsley-Thompson 2001; Deslippe et al. 2001). Knowledge of Tenebrionidae in the transitional coastal desert is limited to the reports by Cepeda-Pizarro et al. (2005), who documented the presence of 20 species belonging to 14 genera for the northern area between 27 and 30° S. Alfaro et al. (2009) documented the presence of 14 species in Pingüino de Humboldt National Reserve,

arranged in eight genera and six tribes, from which seven species were common to the archipelago and five genera were reported for the first time as occurring in insular habiislands: Psectrascelis tat Entomochilus Solier, Diastoleus Solier, Scotobius Germar, and Thinobatis Eschscholtz. Gyriosomus granulipennis Pizarro-Araya & Flores was recorded as endemic to the Choros Island (Pizarro-Araya and Flores 2004; Alfaro et al. 2009; Pizarro-Araya et al. 2012a) and Praocis (Praocis) spinolai Gay & Solier was the most abundant species among the beetles registered at the three islands (Alfaro et al. 2009). Because these islands represent one peculiar insular ecological unit within the transitional coastal desert, the aim of this study was to evaluate the island effect of isolated geographic areas on the morphological differentiation between four populations of *Praocis* (*Praocis*) spinolai using a geometric morphometrics approach.

Materials and Methods

Study area

The study was conducted in the coastal area of Punta de Choros (29° 15' S, 71° 26' W) and Los Choros Archipelago (29° 32' S, 67° 61' W), which includes three islands: Choros (29° 15' S, 71° 32' W), with a surface of 322 ha, Damas (29° 13' S, 71° 31' W), with a surface of 56 ha, and Gaviota (29° 15' S, 71° 28' W), with a surface of 182 ha. This coastal desert area is located ~114 km north of La Serena, Coquimbo Region, Chile (Figure 1). The area has a Mediterranean type climate with morning fog (Di Castri and Hajek 1976).

Sampling

The data on taxonomical composition were collected by means of pitfall traps set up in

the continental and insular ecotopes. The traps were placed in four contrasting pedological units for the continental area, and three for each island. The continental zone was represented by the coastal area of Punta de Choros, which is a coastal desert zone. Four environments were selected for this area, namely coastal steppe, coastal dune, coastal wetland, and interior coastal steppe. These environments were characterized by sandy soil scarcely developed and flat scrubland. The island area was represented by three sites for each one of the islands in the archipelago Los Choros. The sites selected for the island system were characterized by cliffs with stony soils (Isla Choros) and sandy soils (Damas and Gaviota) with poor vegetation (Castro and Brignardello 2005). Two plots $(4 \times 5 \text{ m each})$ were established in each ecotope, and 20 pitfall traps were arranged at 1-m intervals in each plot. Each trap consisted of a plastic jar (70.4 mm diameter, 102 mm height) filled to two-thirds capacity with a 3:7 mixture of formalin (10%) and water with detergent. The traps were active for three days during four months (June, August, October, and November) in 2005 and three months (August, October, and December) in 2006. The material collected was retrieved, cleaned, and preserved in alcohol (70%) until processing. Sampled specimens are now stored in the collection of the Laboratorio de Entomología Ecológica at the Universidad de La Serena, La Serena, Chile (LEULS).

Morphometric analysis

A total of 117 selected specimens of *P*. (*P*) *spinolai* were used for the morphometric analyses. Fifty-five individuals were analyzed from the continental ecotope and 58 from the island ecotope (33 Gaviota, 25 Choros, and 4 Damas). The ventral side of each individual was photographed using a

Nikon Coolpix L120 digital camera (14 megapixel, <u>www.nikon.com</u>). Twenty landmarks were digitized (anatomical homologous points) on every picture with TpsDig 2.10 (Rohlf 2006) (Figure 2). All analyses were then run using MorphoJ software version 1.05a (Klingenberg 2011).

Once the Cartesian x-y coordinates were obfor all landmarks, the tained information was extracted with a full Procrustes fit (generalized Procrustes analysis, Rohlf and Slice 1990; Dryden and Mardia 1998), taking into account the object symthe structure. metry of **Procrustes** superimposition removes the information of size, position, and orientation, and standardizes each specimen to unit centroid size (obtained as the square root of the summed squared Euclidean distances from each landmark to the specimen centroid) and provides an estimation of the size of the studied structure (Dryden and Mardia 1998). For studies of object symmetry, reflection is removed by including the original and mirror image of all configurations in the analysis and superimposing all of them simultaneously (Klingenberg et al. 2002).

Shape variation was analyzed in the entire dataset with principal component analysis (PCA) based on the covariance matrix of symmetric and asymmetric components of shape variation. The first one is the average of left and right sides and represents the shape variation component, whereas the asymmetric component represents the individual left-right differences (Klingenberg et al. 2002).

Differences between locations were assessed using canonical variate analysis, a multivariate statistical method used to find the shape characters that best distinguish among multiple groups of specimens. Because of the lack of specimens for the Damas population, the analysis was run only for the other three populations. The results were reported as Mahalanobis distance and Procrustes distances and the respective *p*-values, after a permutation test that runs 10,000 permutations.

Finally, Procrustes ANOVA for size and shape and MANOVA analyses assessed for studies on object symmetry were performed to evaluate if the observed differences in the sample were due to real differences in the populations examined.

Results

The PCA for the symmetric component (invariation) showed differences dividual between the three populations analyzed. The first two PCs accounted for 55.37% (PC1 + PC2 = 33.33% + 22.04%) of the total shape variation and provided a reasonable approximation of the total amount of variation. The other PC components accounted for no more than 12% of the variation each. The PCA analyses for the asymmetry component (leftright asymmetries) showed differences between populations as well. The first two PCs accounted for 52.33% (PC1 + PC2 = 38.84%+ 13.49%) of the total shape variation, and the other PCs accounted for no more than 9% of the variation each. According to PCA, canonical variate analysis showed significant differences in both symmetric and asymmetric components between the three populapopulations examined and after permutation test (10.000 permutation runs) (Table 1. Figure 3). Finally, Procrustes ANOVA for size did not show significant differences between populations (F = 1.37, p < 0.2545). Procrustes ANOVA for shape showed differences between populations (F = 3.05, p < 0.0001)

Table 1. Results of the canonical variate analysis with Mahalanobis and Procrustes distances and the respective *p*-values for the symmetric and asymmetric components of the variation.

Site	C Mahalanobis p-value	G Mahalanobis p-value	C Procustes p-value	G Procustes p-value	C Mahalanobis p-value	G Mahalanobis p-value	C Procustes p-value	G Procustes p-value
	SYMM	SYMM	SYMM	SYMM	ASYMM	ASYMM	ASYMM	ASYMM
G	2.11	-	0.024	-	0.96	-	0.0034	
	<0.0001		0.0003		0.41		0.59	
I	2.57	2.5744	0.019	0.019	1.79	1.76	0.0062	0.0075
	< 0.0001	< 0.0001	0.004	0.0189	< 0.0001	0.0002	0.045	0.033
_				•				

G: Gaviota, I: Choros, C: Mainland

and high differences among individuals emerged (F = 5.79, p < 0.0001). MANOVA tests, for both symmetric and asymmetric components, confirmed these results (Pillai's trace = 1.09, p < 0.0001; Pillai's trace = 0.78, p < 0.0001, respectively).

Discussion

Morphological differences in both individual and populations were found in this study. The populations examined are well-separated, indicating the existence of an evolutionary pattern.

The observed differences could be due to the isolation of the populations and climaticenvironmental effects, as the islands are subjected to more arid conditions than the continental area, as the continent has a more stable environment due to the presence of coastal wetlands and the coastal range of enabling fog condensation mountains. (Cepeda-Pizarro et al. 2005; Valdivia et al. 2011). Under stochastic processes and environmental stress, the isolated and small populations suffer more than the large interconnected populations, as studies have shown the populations more affected by losses of genetic variability are small and isolated populations (Frankham et al. 2001; Allendorf and Luikart 2007). These losses are often accompanied by a negative impact on individual fitness (Reed and Frankham 2003). In general, the reductions in viability are reflected in morphologic traits or asymmetry. The results presented here indicate

that morphological variations and the variation among sampling sites were mainly due to differences in shape. It is frequently suggested that morphological variation of individuals may be strongly dependent upon unfavorable environmental conditions (Adams and Funk 1997; Tatsuta et al. 2001). In fact, individuals under environmental noise could develop any kind of asymmetries (Van Valen 1962).

Although the differences in body shape observed were not obvious, individuals from the mainland had thinner bodies than those from the islands. It has been reported that a climate with high relative humidity and constant temperatures promotes a thinner subelytral cavity, thus this result was expected for the mainland (Draney 1993; Duncan 2002, 2003).

The individuals of the different islands had more pronounced morphological variation, which may be a consequence of the heterogeneity of the environment in this area (higher variation in temperature ranges, which leads to thicker subelytral cavities). Regarding habitat heterogeneity, Fattorini (2009) analyzed the diversity of Tenebrionidae in contrasting ecotopes in the Mediter-Mediterranean island of Santorini (Greece) and concluded that differences in the composition of tenebrionid assembly could be attributed to climate and substrate type, indicating that these are the most important factors regulating the species diversity.

Observations in other latitudes (Lute desert, Central Iran) have found that morphological variations (e.g., pronotum size) in disjuncpopulations of psammophilic tive Tenebrionidae could be related to factors such as temperature and food availability (Taravati et al. 2009). Zachariassen et al. (1987) showed that Tenebrionidae exhibited the lowest water loss rate compared to other desert insects. The authors proposed that these Tenebrionidae use three major physiological characteristics to conserve water: reduction of cuticular water permeability, reduction of breathing water loss due to subelytral cavity, and reduction of metabolic rate.

Due to the isolation that affects the populations examined, the gene flow has been interrupted between them, and the group shows a particularly high plasticity in the capacity to withstand differences and environmental pressures imposed in each particular environment (Palmer 2000). This capacity was reflected in the high morphological plasticity that emerged and indicates that the populations are evolving.

Acknowledgments

We are grateful to Ivan Benoit (Corporación Nacional Forestal del Gobierno de Chile, CONAF) for assistance with obtaining permits to collect in Pingüino de Humboldt National Reserve, and to CONAF authorities for issuing the permits. Fieldwork was financially supported by FPA 04-015-2006 (CONAMA, Coquimbo Region, Chile) and DIULS-PF13121, VACDDI001 of the Universidad de La Serena to J. Pizarro-Araya.

References

Adams DC, Funk DJ. 1997. Morphometric inferences on sibling species and sexual dimorphism in *Neochlamisus bebbianae* leaf beetles: multivariate applications of the thin-plate spline. *Systematic Biology* 46: 180-194.

Agusto P, Mattoni CI, Pizarro-Araya J, Cepeda-Pizarro J, López-Cortes F. 2006. Comunidades de escorpiones (Arachnida: Scorpiones) del desierto costero transicional de Chile. *Revista Chilena de Historia Natural* 79: 407-421.

Alcayaga OE, Pizarro-Araya J, Alfaro FM, Cepeda-Pizarro J. 2013. Arañas (Arachnida: Araneae) asociadas a agroecosistemas en el Valle de Elqui (Region de Coquimbo, Chile). *Revista Colombiana de Entomología* 39: 150-154.

Alfaro FM, Pizarro-Araya J, Flores GE. 2009. Epigean tenebrionids (Coleoptera: Tenebrionidae) from the Choros Archipelago (Coquimbo Region, Chile). *Entomological News* 120: 125-130.

Alfaro FM, Pizarro-Araya J, Letelier L, Cepeda-Pizarro J. 2013. Patrones distribucionales de ortópteros (Insecta: Orthoptera) de las provincias biogeográficas de Atacama y Coquimbo (Chile). *Revista de Geografia Norte Grande* 56: 235-250.

Alibert P, Moureau B, Dommergues JL, David B. 2001. Differentiation at a microgeographical scale within two species of ground beetle, *Carabus auronitens* and *C. nemoralis* (Coleoptera, Carabidae): a geometrical morphometric approach. *Zoologica Scripta* 30: 299-311.

Allendorf FW, Luikart G. 2007. Conservation and the genetics of populations. Blackwell.

Benítez HA. 2013. Assessment of patterns of fluctuating asymmetry and sexual dimorphism in carabid body shape. *Neotropical Entomology* 42: 164-169.

Benítez HA, Briones R, Jerez V. 2008. Asimetría Fluctuante en dos poblaciones de *Ceroglossus chilensis* (Coleoptera Carabidae) en el agroecosistema *Pinus radiata*, Región del BioBío. *Gayana* 72: 131-139.

Castro C, Brignardello L. 2005. Geomorfología aplicada a la ordenación territorial de litorales arenosos. Orientaciones para la protección, usos y aprovechamiento sustentables del sector de Los Choros, Comuna de La Higuera, IV Región. *Revista de Geografia Norte Grande* 33: 33-58.

Clarke GM. 1993. The genetic basis of developmental stability. I. Relationships between stability, heterozygosity and genomic coadaptation. *Genetica* 89: 15-23.

Cloudsley-Thompson JL. 2001. Thermal and water relations of desert beetles. *Naturwissenschaften* 88: 447-460.

Cepeda-Pizarro J, Vega S, Vásquez H, Elgueta M. 2003. Morfometría y dimorfismo sexual de *Elasmoderus wagenknechti* (Liebermann) (Orthoptera: Tristiridae) en dos eventos de irrupción poblacional. *Revista Chilena de Historia Natural* 76: 417-435.

Cepeda-Pizarro J, Pizarro-Araya J, Vásquez H. 2005. Variación en la abundancia de

Arthropoda en un transecto latitudinal del desierto costero transicional de Chile, con énfasis en los tenebriónidos epígeos. *Revista Chilena de Historia Natural* 78: 651-663.

Deslippe RJ, Salazar JR, Guo YL. 2001. A darkling beetle population in West Texas during the 1997-1998 El Niño. *Journal of Arid Environments* 49: 711-721.

Di Castri F, Hajek ER. 1976. *Bioclimatología de Chile*. Imprenta-Editorial de la Universidad Católica de Chile, Santiago, Chile.

Draney ML. 1993. The subelytral cavity of desert tenebrionids. *Florida Entomologist* 76: 539-549.

Dryden IL, Mardia KV. 1998. *Statistical Shape Analysis*. Wiley.

Duncan FD. 2002. The role of the subelytral cavity in water loss in the flightless dung beetle *Circellium bacchus* (Coleoptera: Scarabaeinae). *European Journal of Entomology* 99: 253-258.

Duncan FD. 2003. The role of the subelytral cavity in respiration in a tenebrionid beetle, *Onymacris multistriata* (Tenebrionidae: Adesmiini). *Journal of Insect Physiology* 49: 339-346.

Emerson BC. 2008. Speciation on islands: what are we learning? *Biological Journal of the Linnean Society* 95: 47-52.

Fattorini S. 2009. Darkling beetle communities in two geologically contrasting biotopes: testing biodiversity patterns by microsite comparisons. *Biological Journal of the Linnean Society* 98: 787-793.

Floate KD, Fox AS. 2000. Flies under stress: a test of fluctuating asymmetry as a biomonitor of environmental quality. *Ecological Applications* 10: 1541-1550.

Frankham R, Gilligan DM, Morris D, Briscoe DA. 2001. Inbreeding and extinction: Effects of purging. *Conservation Genetics* 2: 279-285.

Grant PR. 1998. *Evolution on islands*. Oxford University Press.

Jerez V. 2000. Diversidad y patrones de distribución geográfica de insectos coleópteros en ecosistemas desérticos de la región de Antofagasta, Chile. *Revista Chilena de Historia Natural* 73: 79-92.

Klingenberg CP. 2011. MORPHOJ: an integrated software package for geometric morphometrics. *Molecular Ecology Resources* 11: 353-357.

Klingenberg CP, Barluenga M, Meyer A. 2002. Shape analysis of symmetric structures: quantifying variation among individuals and asymmetry. *Evolution* 56: 1909-1920.

Laborda A, Ramírez MJ, Pizarro-Araya J. 2013. New species of the spider genera *Aysenia* and *Aysenoides* from Chile and Argentina: description and phylogenetic relationships (Araneae: Anyphaenidae, Amaurobioidinae). *Zootaxa* 3731(1): 133-152.

Losos JB, Ricklefs RE. 2009. Adaptation and diversification on islands. *Nature* 457: 830-836.

Mayr E. 1967. The challenge of island faunas. *Australian Natural History* 15: 369-374.

Mila B, Warren B, Heeb P, Thebaud C. 2010. The geographic scale of diversification on islands: genetic and morphological divergence at a very small spatial scale in the Mascarene grey white-eye (Aves: *Zosterops borbonicus*). *BMC Evolutionary Biology* 10: 158.

Ojanguren-Affilastro AA, Pizarro-Araya J. In press. Two new scorpion species from Paposo, in the Coastal desert of Taltal, Chile (Scorpiones, Bothriuridae, *Brachistosternus*). *Zootaxa*.

Ojanguren-Affilastro AA, Agusto P, Pizarro-Araya J, Mattoni CI. 2007. Two new scorpion species of genus *Brachistosternus* (Scorpiones: Bothriuridae) from northern Chile. *Zootaxa* 1623: 55-68.

Palmer M 2002. Testing the 'island rule' for a tenebrionid beetle (Coleoptera, Tenebrionidae). *Acta Oecologica-International Journal of Ecology* 23: 103-107.

Piscart C, Moreteau JC, Beisel JN. 2005. Decrease of fluctuating asymmetry during ontogeny in an aquatic holometabolous insect. *Comptes Rendus Biologies* 328: 912-917.

Pizarro-Araya J, Flores GE. 2004. Two new species of *Gyriosomus* Guérin-Méneville from Chilean coastal desert (Coleoptera: Tenebrionidae: Nycteliini). *Journal of the New York Entomological Society* 112: 121-126.

Pizarro-Araya J, Jerez V. 2004. Distribución geográfica del género *Gyriosomus* Guérin-Méneville, 1834 (Coleoptera: Tenebrionidae): una aproximación biogeográfica. *Revista Chilena de Historia Natural* 77: 491-500.

Pizarro-Araya J, Cepeda-Pizarro J, Flores GE. 2008. Diversidad taxonómica de los artrópodos epígeos de la Región de Atacama (Chile): estado del conocimiento. In: Squeo FA, Arancio G, Gutiérrez JR, Editors. *Libro Rojo de la Flora Nativa y de los Sitios Prioritarios para su Conservación: Región de Atacama*. pp. 257-274. Ediciones Universidad de La Serena, La Serena, Chile.

Pizarro-Araya J, Vergara OE, Flores GE. 2012a. *Gyriosomus granulipennis* Pizarro-Araya & Flores 2004 (Coleoptera: Tenebrionidae): Un caso extremo a conservar. *Revista Chilena de Historia Natural* 85: 345-349.

Pizarro-Araya J, Alfaro FM, Castillo JP, Ojanguren-Affilastro AA, Agusto P, Cepeda-Pizarro J. 2012b. Assemblage of arthropods in the Quebrada del Morel private protected area (Atacama Region, Chile). *Pan-Pacific Entomologist* 88: 1-14.

Reed DH, Frankham R. 2003. Correlation between fitness and genetic diversity. *Conservation Biology* 17: 230-237.

Rettig JE, Fuller RC, Corbett AL, Getty T. 1997. Fluctuating asymmetry indicates levels of competition in an even-aged poplar clone. *Oikos* 80: 123-127.

Rohlf FJ. 2006. *TpsDig V2.10*. Department of Ecology and Evolution, State University of New York. Available online: http://life.bio.sunysb.edu/morph/

Rohlf FJ, Slice D. 1990. Extensions of the Procrustes method for the optimal superimposition of landmarks. *Systematic Zoology* 39: 40-59.

Taravati S, Darvish J, Mirshamshi O. 2009. Geometric morphometric study of two species of the psammophilous genus *Erodiontes* (Coleoptera: Tenebrionidae) from the Lute desert, Central Iran. *Iranian Journal of Animal Biosystematics* 5: 81-89.

Tatsuta H, Mizota K, Akimoto SI. 2001. Allometric patterns of heads and genitalia in the stag beetle *Lucanus maculifemoratus* (Coleoptera: Lucanidae). *Annals of the Entomological Society of America* 94: 462-466.

Valdivia DE, Pizarro-Araya J, Cepeda-Pizarro J, Ojanguren-Affilastro AA. 2008. Diversidad taxonómica y denso-actividad de solífugos (Arachnida: Solifugae) asociados a un ecosistema desértico costero del centro norte de Chile. *Revista de la Sociedad Entomológica Argentina* 67: 1-10.

Valdivia DE, Pizarro-Araya J, Briones R, Ojanguren-Affilastro AA, Cepeda-Pizarro J. 2011. Taxonomical diversity and abundance of solpugids (Arachnida: Solifugae) in coastal ecotopes of north-central Chile. *Revista Mexicana de Biodiversidad* 82: 1234-1242.

Van Valen L. 1962. A study of fluctuating asymmetry. *Evolution* 16: 125-142.

Zachariassen KE, Andersen J, Maloiy GMO, Kamau JMZ. 1987. Transpiratory water loss and metabolism of beetles from arid areas in East Africa. *Comparative Biochemistry and Physiology A*. 86: 403-408.

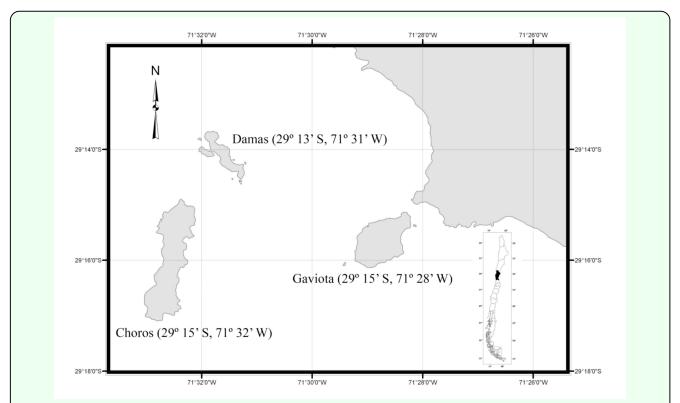


Figure 1. Map of the mainland of Los Choros indicating the study area and the sampling sites, the islands of Choros, Damas, and Gaviota (Coquimbo Region, Chile). High quality figures are available online.

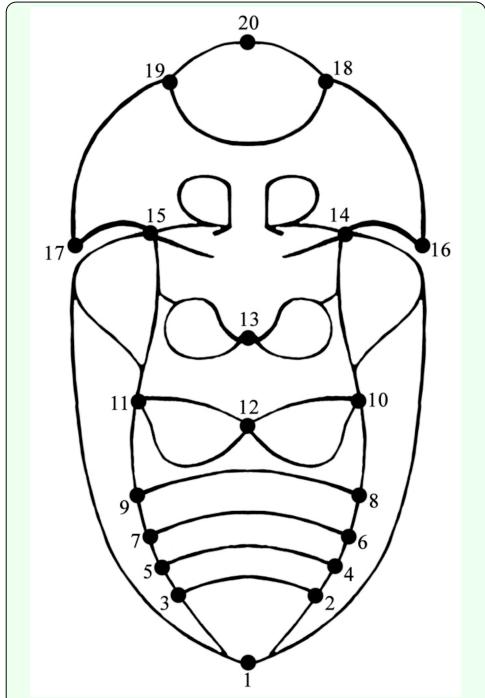


Figure 2. Indication of 20 landmarks in the ventral view of *Praocis* (*Praocis*) *spinolai*. 1: pygidium, 2: right lateral vertex of abdominal segment 4, 3: left lateral vertex of abdominal segment 4, 4: right lateral vertex of abdominal segment 3, 5: left lateral vertex of abdominal segment 3, 6: right lateral vertex of abdominal segment 2, 7: left lateral vertex of abdominal segment 2, 8: right lateral vertex of abdominal segment 1, 9: left lateral vertex of abdominal segment 1, 10: right lateral vertex of metastern, 11: left lateral vertex of metastern, 12: mean point of metastern, 13: lower mean point of mesostern, 14: right vertex of pronotal epimere, 15: left vertex of pronotal epimere, 16: right pronotal posterior angle, 17: left pronotal posterior angle, 18: right vertex of lip, 19: left vertex of lip, 20: mean point of head between mandibles. High quality figures are available online.

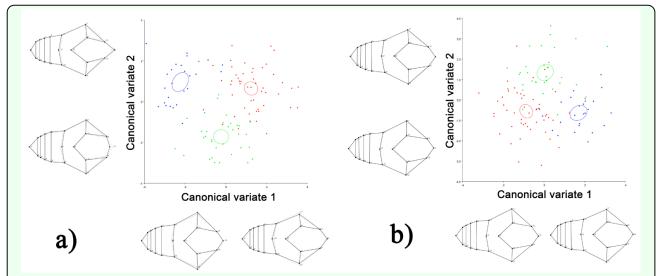


Figure 3. Canonical variate analysis of three of the four isolated populations of *Praocis* (*Praocis*) *spinolai*: Gaviota (green), Choros (blue), and mainland (red). In the figure are shown the first two canonical variate compenents' axes with shape deformation images associated. (A) Canonical variate analysis for the symmetric component, (B) canonical variate analysis for the asymmetric component. High quality figures are available online.